

## VERAISON LEAF REMOVAL MODIFY ANTHOCYANIN AND FLAVONOL PROFILE IN FOUR *Vitis Vinifera* L. CULTIVARS.

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### Introduction

Defoliation is a common crop management practice on grapevine in many viticultural regions. The elimination of a certain number of basal leaves conventionally applied in the fruiting zone from berry set to veraison, enhances air circulation, berries sunlight exposure and increases berry temperature, while reduces Botrytis bunch rot infection and increases fungicide spray penetration (English et al., 1989; Stapleton and Grant, 1992; Zoecklein et al., 1992).

Especially the effects of veraison defoliation on grape composition have been shown to be strongly influenced by intensity of treatment, genotype and climatic conditions (Downey et al., 2006; Guidoni et al., 2008; Hunter et al., 1991, Matus et al., 2009).

Leaf removal applied at veraison has a strong impact on bunch microclimate and a limited impact on the vine source–sink balance due to the lower photosynthetic activity of basal leaves compared to the intermediate and apical leaves at that stage (Poni et al., 1994). In general, after leaf removal, bunches are subjected to synergistic effects due to increase of light and temperature that, depending on the seasonal and climatic conditions, may affect grape composition. Several authors, mainly reporting the effects of shading on grape color, agreed that low light reduces anthocyanin and other flavonoid concentrations, while increasing light increases the flavonoid content of grapes (Crippen and Morrison, 1986 a, b; Dokoozlian and Kliewer, 1996; Hale and Buttrose, 1974; Hunter et al., 1991; Iland, 1988; Matus et al., 2009; Zoecklein et al., 1992).

Further investigations into the effects of increasing light exposure on grape color gave rise to contradictory results. Some studies reported that high light levels resulted in decreased anthocyanin levels (Bergqvist et al., 2001; Pastore et al., 2013; Spayd et al., 2002), while in other cases no change was observed in total anthocyanin concentration (Downey et al., 2004; Haselgrove et al., 2000; Price et al., 1995).

When exposure to sunlight is associated with excessive berry temperature, as occurs in warm conditions, this may often lead to berry sunburn that has a negative impact on the color of some red (Kliewer and Torres, 1972; Mori et al., 2005; Mori et al., 2007) and white berry grapevine varieties also due to photo-oxidation (Rustioni et al., 2015). It has been pointed out that the lower anthocyanin content in berries under high temperature reflects the combined impact of reduced biosynthesis and increased degradation in which the role of peroxidase enzymes in anthocyanin catabolism is probably involved (Movahed et al., 2016).

The modification of bunch light exposure around veraison can also affect anthocyanin composition. As is well-known, grape anthocyanins are based on cyanidin, peonidin, delphinidin, petunidin and malvidin that are glycosylated at the third position of the C ring. Several researches have shown shifts

in anthocyanin composition after bunches microclimatic variation, with an increase in the di-substituted anthocyanin concentration (cyanidin and peonidin) in shaded bunches giving rise to an increased di-substituted to tri-substituted anthocyanins (delphinidin, petunidin and malvidin) ratio (Downey et al., 2004; Ristic et al., 2007; Spayd et al., 2002). Other authors showed opposite results, since bunch light exposure increased the proportion of di- respect to tri-substituted anthocyanins (Chorty et al., 2010; Guidoni et al., 2008; Tarara et al., 2008).

These contradictory outcomes also in terms of composition may be probably ascribed again to both light and temperature effects, which frequently coexist, playing a conflicting role especially in warm climatic conditions. Sunlight is known to enhance flavonol accumulation in berries (Downey et al., 2006) and several papers focused on the effects of solar UV radiation, suggest a strong positive correlation between illumination and flavonol levels, reflecting their role as UV protectants (Carbonell-Bejerano et al., 2014; Price et al., 1995; Spayd et al., 2002). High accumulation of flavonols was also observed in different varieties subjected to leaf removal compared to controls (Lemut et al., 2013; Pereira et al., 2006) and this was also supported by an increase in flavonol synthase gene expression in the berries (Pastore et al., 2013).

Although in Sangiovese berries a shift in flavonol composition was registered after veraison defoliation due to higher accumulation of quercetin and kaempferol than myricetin compared to control berries (Pastore et al., 2013), studies on other cultivars have shown that the abundance of all flavonol compounds increases with the same intensity following defoliation (Spayd et al., 2002).

Considering that the profile of anthocyanins (Mattivi et al., 2006) and flavonols (Downey et al., 2003) in each variety are relatively stable over seasons and that distinctive varietal responses to light and temperature may be observed in flavonol and anthocyanin accumulation and composition in berry skin (Mattivi et al., 2006), the aim of this study was to analyse anthocyanin and flavonol composition of berries at harvest by describing the response of four red varieties, characterized by different anthocyanin and flavonol profiles, to veraison leaf removal over two years.

## Material and methods

The trial was conducted in 2008 and 2009 on adult *Vitis vinifera* L. Cabernet Sauvignon, Nero d'Avola, Raboso Piave and Sangiovese vines grafted to SO4, in a vineyard with no irrigation system located in Bologna, Italy (44°30'N, 11°24'E), with north–south oriented rows. Vine spacing was 1.0 m x 3.0 m and the training system was a vertical shoot positioned spur pruned cordon (12 buds per vine), with cordon height at 1.0 m above the ground and canopy height of about 1.3-1.4 m. Pest management followed local practices in the Emilia Romagna Region. Each vine in the trial was uniformed for bud load and bunch number at flowering. Nine vines per treatment in three blocks were selected in a single uniform row and each vine was randomly assigned to the following treatments: a) control (C), no treatment; b) veraison defoliation (D), hand defoliation of six basal leaves at veraison. In the defoliation treatments, any laterals growing in the 6 basal node of the main shoot were also removed. Defoliation treatments were performed at the beginning of veraison, with sugar concentration around 8 ° Brix and each variety was harvested when the soluble solids concentration was stable for about a week as reported in Table 1.

Weather data (mean daily air temperature and rainfall) were recorded from April to September in both years, by a meteorological station located close to the experimental site.

	2008		2009	
	Defoliation	Harvest	Defoliation	Harvest
<b>Cabernet Sauvignon</b>	226 (13th August)	276 (2th October)	217 (5th August)	271 (28th September)
<b>Nero d'Avola</b>	226 (13th August)	276 (2th October)	217 (5th August)	271 (28th September)
<b>Raboso Piave</b>	226 (13th August)	287 (13th October)	225 (13th August)	281 (8th October)
<b>Sangiovese</b>	211 (29th July)	266 (22th September)	210 (29th July)	261 (18th September)

Table 1. Dates (DOY, days and months) on which veraison defoliation treatment and harvest took place in 2008 and 2009 for Cabernet Sauvignon, Nero d'Avola, Raboso Piave and Sangiovese.

### Agronomic parameters at harvest

At harvest the number and weight of bunches per vine were measured. For each bunch we determined the sunburn and Botrytis incidence according the guide line EPPO PP 1/17 (<http://pp1.eppo.int>) with the evaluation of the percentage of surface area affected for each bunch at harvest. During winter, the wood pruned from each vine was weighed.

### Temperature monitoring

Berry skin temperature was monitored in 2008 and 2009 in four selected bunches on control and defoliated vines of each tested variety. For each treatment, temperature data were collected from stage 33 (beginning of bunch closure, berries touching, according to Lorenz et al., 1995) until harvest and this fluctuated for each cv: Cabernet Sauvignon and Nero d'Avola from DOY 226 to 276 in 2008 and from DOY 217 to 271 in 2009; Raboso Piave from DOY 226 to 287 in 2008 and from DOY 225 to 281 in 2009; Sangiovese from DOY 211 to 265 in 2008 and from DOY 210 to 261 in 2009. Eight T-type thermocouples (RS components, MI, Italy) were positioned in the sub-cuticular tissues of the berry skin. Four were positioned on two different bunches, two on the east side and two on the west side of the cordon. For each side, one thermocouple was inserted in a berry located in the external part of the bunch and the other in the internal part. Each probe was then connected to a CR10X data logger (Campbell Scientific Ltd., Leicestershire, UK) that registered temperature data every 15 minutes. In three days during August in 2008 and in 2009 for each bunch, the percentage of bunch exposure was visually estimated in three moments of the day: in the morning (9.00-9.30 a.m.), when the sun position is at its Zenith (1.30- 2.00 p.m.) and in late afternoon (5.30-6.00 p.m.).

### Biochemical analysis

For each treatment, we collected 40 berries from each of the three vines in each block at harvest. The samples were divided into two parts. Twenty berries were weighed and immediately tested for ripening by crushing and filtering the must through a strainer for the evaluation of °Brix, titratable acidity and pH. The anthocyanins and flavonols were extracted from the skins of the 20 remaining berries by soaking the peeled skins in 100 mL methanol for 24 h, then storing the extracts at -20°C (Mattivi et

al., 2006). To analyze the concentration of each flavonol aglycone the acid hydrolyzation of flavonols glucoside was conducted (Mattivi et al., 2006). Anthocyanins and flavonols were separated by HPLC using a Waters 1525 instrument equipped with a diode array detector (DAD) and a reversed-phase column (RP18 250 x 4 mm, 5  $\mu$ M) with a pre-column (Phenomenex, Castel Maggiore, BO, Italy). Anthocyanins were quantified at 520 nm using an external calibration curve with malvidin-3-glucoside chloride as the standard (Sigma-Aldrich). Flavonols were quantified at 370 nm with the corresponding external standards (myricetin, quercetin and kaempferol) purchased from Extrasynthese (Genay, France).

### **Statistical analyses**

Yield components and grape composition parameters were processed for each variety by analysis of variance using the mixed procedure available in SAS v9.0 (SAS Institute, Inc., Cary, NC, USA). Treatment comparisons were analyzed using the Tukey test with a cut-off at  $P \leq 0.05$ . To compare anthocyanin and flavonol composition in different varieties, treatments and years, multivariate analysis was applied on the data of each compound. An exploratory principal component analysis was performed separately on anthocyanins and flavonols to point out differences and any gradients.

## **Results and discussion**

### **Climatic data and impact of defoliation on berry skin temperature and vegetative and productive traits**

The weather during 2008 and 2009 was on the average of the area and total rainfall from April through September was very similar in the two seasons (320 mm and 317.4 mm respectively). Mean and maximum temperature (Figure 1) during the growing season in 2008 (19.8 °C and 35.9 °C respectively) was lower than in 2009 (20.9 °C and 36.8° C respectively) and this reflected on total active heat summation calculated using base 10 °C days from April through September (1758 °C in 2008 and 2006 °C in 2009).

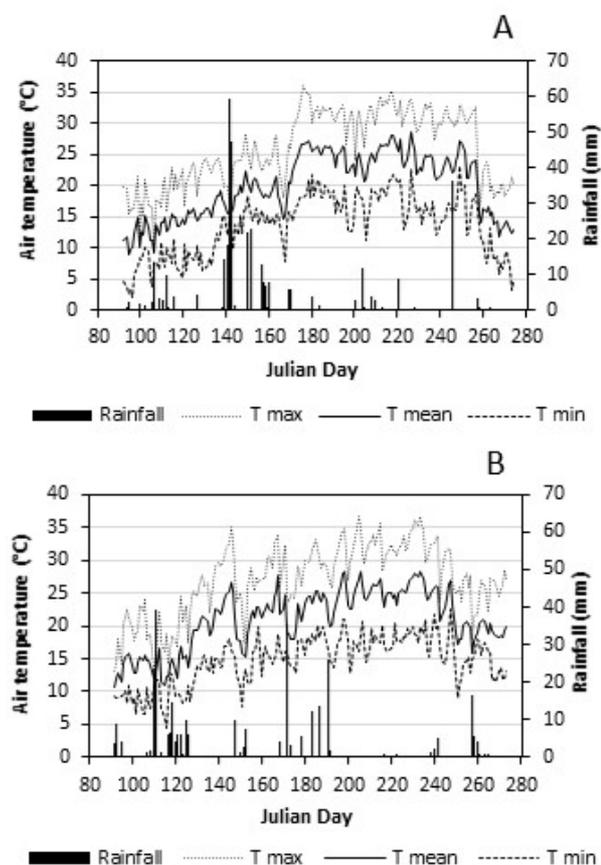


Figure 1. Seasonal trends (1 April–30 September) of diurnal air mean, maximum and minimum temperature recorded close to the trial site in (A) 2008 and (B) 2009. Vertical bars indicate daily rainfall. The Degree Days and total rainfall from 1 April to 30 September were, respectively, 1768 and 332 mm in 2008 and 2006 and 317 mm in 2009.

Sangiovese was the earliest variety for both veraison and harvest, while Raboso Piave was the latest. It should be noticed that the number of days between veraison and harvest was similar among varieties and ranged from 50 to 61.

We monitored the berry skin temperature from the application of leaf removal until harvest in the control and defoliated vines of each variety. The berries of all tested varieties in the control treatment were exposed to temperatures >30 °C for less time than in the defoliated samples with differences between the two treatments ranging from up to 70 hours to a minimum of 31 hours (for the same cv Sangiovese respectively in 2009 and 2008, Table 2). In both treatments, the number of hours with berry temperature above 30 °C was higher in 2009 than in 2008. The estimation of the percentage of bunch exposure after defoliation showed in both years an increase of around 20 % in the daily average (Table 2). There were only minor differences between the two years in vegetative and productive measurements at harvest following the leaf removal in all tested varieties. Starting from a uniform bunch number per vine, no differences were detected after defoliation in yield per vine or berry mass at harvest, for either variety or year (Table 3). Raboso Piave and Sangiovese showed a significant increase in the percentage of sunburned bunches on defoliated compared with control vines, whereas

it should be noted that the untreated Nero d'Avola was the most sensitive cultivar to Botrytis, showing the highest level of attack.

Parameter	2008		2009	
	C	D	C	D
<b>Cabernet Sauvignon</b>				
h>30 °C	147 b	202 a	214 b	270 a
Average bunch exposure (%)	5.2 b	24.8 a	6.2 b	26.4 a
<b>Nero d'Avola</b>				
h>30 °C	145 b	205 a	212 b	263 a
Average bunch exposure (%)	3.3 b	23.4 a	4.2 b	25.3 a
<b>Raboso Piave</b>				
h>30 °C	147 b	202 a	164 b	206 a
Average bunch exposure (%)	2.1 b	23.8 a	3.2 b	24.6 a
<b>Sangiovese</b>				
h>30 °C	269 b	300 a	256 b	324 a
Average bunch exposure (%)	5.4 b	26.4 a	6.7 b	26.3 a

Table 2. Number of hours during which berry temperature was higher than 30 °C on control (C) and defoliated (D) vines during the experimental period. For each variety and year, the period of measurements ranges from leaf removal to harvest and are as follows: Cabernet Sauvignon and Nero d'Avola from DOY 226 to 276 in 2008 and from DOY 217 to 271 in 2009; Raboso Piave from DOY 226 to 287 in 2008 and from DOY 225 to 281 in 2009; Sangiovese from DOY 211 to 265 in 2008 and from DOY 210 to 261 in 2009. Values represent means of eight replicates. Average of percentage of bunch exposure estimated in 2008 and 2009. For each variety and year, the measurements were performed in three days during August at 9.00 am, 1.30 pm and 5.30 pm.

Surprisingly, Sangiovese cv, despite a strong Botrytis incidence, did not respond to leaf removal with significant rot reduction (Table 3). Sugar concentration in must at harvest was not affected by veraison defoliation, while total acidity and pH in Cabernet Sauvignon, Nero d'Avola and Sangiovese were reduced and increased respectively by defoliation (Table 3).

Parameter		
	C	D
<b>Cabernet Sauvignon</b>		
Yield /vine (kg)	3.09	3.54
Berry mass (g)	1.48	1.50
Botrytis (%)	2.50	0.00
Sunburn (%)	0.00	0.00
Total Soluble Solids (°Brix)	22.16	21.77
Titrateable acidity (g/L)	7.07 a	5.91 b
pH	3.61 b	3.70 a
<b>Nero d'Avola</b>		
Yield /vine (kg)	4.51	4.17
Berry mass (g)	2.40	2.31
Botrytis (%)	9.50 a	1.70 b
Sunburn (%)	0.00	1.00
Total Soluble Solids (° Brix)	22.07	21.89
Titrateable acidity (g/L)	7.79 a	7.06 b
pH	3.36 b	3.42 a
<b>Raboso Piave</b>		
Yield /vine (kg)	4.20	3.24
Berry mass (g)	1.99 a	1.87 b
Botrytis (%)	0.50	0.00
Sunburn (%)	1.10 b	18.50 a
Total Soluble Solids (° Brix)	22.34	21.88
Titrateable acidity (g/L)	11.21	11.65
pH	3.21	3.23
<b>Sangiovese</b>		
Yield /vine (kg)	6.71	5.72
Berry mass (g)	2.51	2.42
Botrytis (%)	9.70	6.95
Sunburn (%)	0.75 b	9.55 a
Total Soluble Solids (° Brix)	20.89	21.42
Titrateable acidity (g/L)	7.28a	6.42b
pH	3.41b	3.49a

Table 3. Yield components and main grape composition parameters recorded at harvest in Cabernet Sauvignon, Nero d'Avola, Raboso Piave and Sangiovese vines subjected to defoliation at veraison (D) in comparison to control vines (C). Data are means over 2008-2009. Different letters within row indicate significant differences between the treatments by Tukey test ( $P < 0.05$ ). No year x treatment interaction were registered. Botrytis and sunburn incidence were expressed as average percentage of surface area with symptoms for each bunch at harvest.

## Anthocyanins and flavonols

### Univariate analyses

The concentration of total anthocyanins in the berries (mg/g) did not vary among treatments at harvest in both vintages and in all varieties (Table 4). The di-substituted to tri-substituted anthocyanins ratio significantly increased with defoliation in Nero d' Avola and Sangiovese cultivars. Raboso Piave showed a similar tendency but without significant differences between treatments. The concentration of total flavonols at harvest increased significantly in defoliated berries of all varieties compared to controls in both years (Table 5). Each variety showed a characteristic composition in control berries as quercetin is the main component in Sangiovese, myricetin is in Nero d'Avola, while Raboso Piave and Cabernet Sauvignon showed similar proportions of quercetin and myricetin. The total flavonols increase was quite similar in all varieties but each flavonol compound showed a different increment following leaf removal. The highest proportional increase concerned quercetin in Raboso Piave (Table 5).

	C	D
<b>Cabernet Sauvignon</b>		
Total anthocyanins	5.96	5.33
Di-Tri substituted ratio	0.205 a	0.117 b
<b>Nero d'Avola</b>		
Total anthocyanins	8.42	8.51
Di-Tri substituted ratio	0.068 b	0.101 a
<b>Raboso Piave</b>		
Total anthocyanins	9.42	9.68
Di-Tri substituted ratio	1.209	1.347
<b>Sangiovese</b>		
Total anthocyanins	4.58	4.52
Di-Tri substituted ratio	0.830 b	1.456 a

Table. 4. Concentration of total anthocyanins (mg/g skin) and ratio between di-substituted and tri-substituted anthocyanins at harvest in Cabernet Sauvignon, Nero d'Avola, Raboso Piave and Sangiovese vines subjected to defoliation at veraison (D) in comparison to control vines (C). Data are means over 2008-2009. Different letters within row indicate significant differences between the treatments by Tukey test ( $P < 0.05$ ). No year x treatment interaction were registered.

	C	D
<b>Cabernet Sauvignon</b>		
Total flavonols	0.20 b	0.48 a
Myricetin	0.10 b	0.21 a
Quercetin	0.09 b	0.23 a
Kaempferol	0.01 b	0.04 a
<b>Nero d'Avola</b>		
Total flavonols	0.32 b	0.77 a
Myricetin	0.20 b	0.38 a
Quercetin	0.11 b	0.33 a
Kaempferol	0.01 b	0.06 a
<b>Raboso Piave</b>		
Total flavonols	0.14 b	0.49 a
Myricetin	0.07 b	0.11 a
Quercetin	0.07 b	0.36 a
Kaempferol	0.00 b	0.03 a
<b>Sangiovese</b>		
Total flavonols	0.36 b	0.68 a
Myricetin	0.06 b	0.07 a
Quercetin	0.28 b	0.56 a
Kaempferol	0.02 b	0.05 a

Table. 5. Concentration of total and single flavonol compounds (mg/g skin) at harvest in Cabernet Sauvignon, Nero d'Avola, Raboso Piave and Sangiovese vines subjected to defoliation at veraison (D) in comparison to control vines (C). Data are means over 2008-2009. Different letters within row indicate significant differences between the treatments by Tukey test ( $P < 0.05$ ). No year x treatment interaction were registered.

### Multivariate quantitative data

Comprehensive analysis of the total dataset of anthocyanin (Figure 2) and flavonol (Figure 3) concentrations in mg per gram of berry skin of the varieties Cabernet Sauvignon, Nero D'Avola, Raboso Piave and Sangiovese in 2008 and 2009, was conducted, applying an exploratory principal component analysis separately on anthocyanins and flavonols to evaluate the distribution of single observations and rank the data.

As presented in Fig. 2, 90% of the variability due to anthocyanin concentration is accounted for the two discriminant functions. The first one accounts for 55% of the information and is mainly correlated with the concentration of cyanidin 3-glucoside and peonidin 3-glucoside on one side and malvidin 3-glucoside on the other. Sangiovese and Raboso Piave are close to each other and clearly separated from Nero d'Avola, which is near Cabernet Sauvignon, according to the first component (PC1), by bunching at positive and negative PC1 values, respectively (Figure 1). The second function (PC2) accounts for 35% of the variability and seems to be responsible for the differences between treatments and years. Raboso Piave shows high variability and treatments are not clearly separated, while it is possible to identify a separation in Sangiovese between defoliated and control vines

independently of the season. In Cabernet Sauvignon, the two years appear grouped and in Nero d'Avola the two treatments are distinguished mainly according to the second component (PC2).

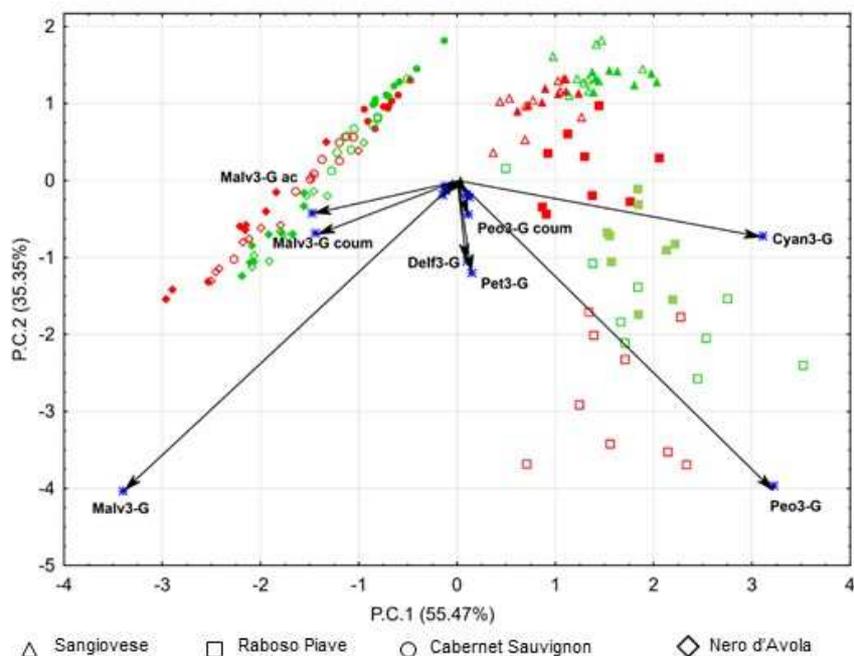


Figure 2. Principal component analysis of the total data set of anthocyanin concentrations (mg per gram of berry skin) of control (red) and defoliated (green) of Cabernet Sauvignon, Nero D'Avola, Raboso Piave and Sangiovese in 2008 (empty) and 2009 (full). The name of single anthocyanin compound responsible of cultivars, treatments and seasons scattering, are represented with arrows and asterisks. In particular, each name corresponds to: Malv-3-G, malvidin 3-glucoside; Malv3-G ac, malvidin-3-acetyl-glucoside; Malv 3-G coum, malvidin 3-coumaroyl glucoside; Del 3-G, delphinidin 3-Glucoside; Peo3-G, peonidin 3-glucoside; Peo3-G coum, peonidin 3-coumaroyl glucoside; Cyan 3-G, cyanidin 3-Glucoside.

The same approach was applied for flavonol concentration and the results are reported in Figure 3 where the two discriminant functions account for more than 99% of the variability. The PC1 accounts for 70.9% of the variability mainly linked to the variation in quercetin. For all varieties, it is possible to separate the control from defoliated vines according to the PC1.

The second function (PC2), which accounts for 28.8% of the variability, is dependent mainly on myricetin. According to this function, the observations allow genotype separation with Nero d'Avola and Cabernet Sauvignon mainly matched with positive values, while Sangiovese and Raboso Piave with the negative values of PC2 (Figure 3).

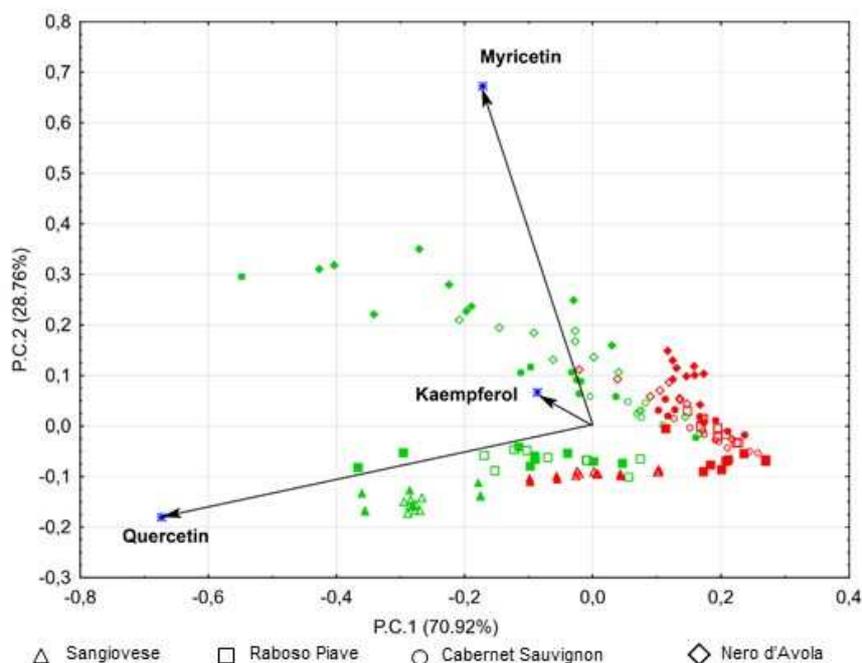


Figure 3. Principal component analysis of the total data set of flavonols concentrations (mg per gram of berry skin) of control (red) and defoliated (green) of Cabernet Sauvignon, Nero D'Avola, Raboso Piave and Sangiovese in 2008 (empty) and 2009 (full). The name of single flavonol compound (myricetin, kaempferol and quercetin) responsible of cultivars, treatments and seasons scattering, are represented with arrows and asterisks.

## Discussion

### Vegetative and productive traits

The four varieties included in this research, Sangiovese, Cabernet Sauvignon, Nero d'Avola and Raboso Piave, as expected did not modify vegetative and yield traits as a result of veraison leaf removal. In fact, veraison defoliation, with the elimination of already senescent basal leaves, may have a limited effect on the vine source-sink balance and on berries sugar accumulation (Bledsoe et al., 1988; Pastore et al., 2013; Percival et al., 1994).

On the other hand, veraison defoliation usually had strong impact on bunches microclimatic conditions. In our study, we estimated an average daily increase of 20% of bunch exposure in defoliated compared to control vines, in both years, while the berry temperature difference between the treatments within all cultivars and years, expressed as number of hours in which the berries overcome 30°C from veraison to harvest, never exceeded 70 hours. As well known in grapevines at temperature overcoming 30° C many metabolic processes stop or are significantly reduced, as reviewed by Downey et al., (2006), whereas the critical temperature leading to the inhibition of anthocyanin synthesis is reported to be between 30 and 35 °C, varying according to different authors (Coombe, 1986; Kliewer and Torres, 1972; Mohaved et al., 2016; Mori et al., 2005). During the two seasons the maximum air temperature was around 36.5 °C.

Although we did not measure the individual malic and tartaric acid fractions, we could argue that the decrease in total acidity registered following defoliation in three of the four varieties, Cabernet Sauvignon, Nero d'Avola and Sangiovese, independently of sugar concentration, is correlated to the

thermal increase due to higher bunch exposure to light, since light is not known to influence malic and tartaric acid accumulation in grape tissues (Crippen and Morrison, 1986 a; Kliewer and Lider, 1968). The fact that the acidity concentration in Raboso Piave did not decrease as a result of defoliation treatment, suggests a cultivar-dependent thermal response of acidity, as previously reported on different cultivars subjected to increased temperature regime (Bergqvist et al., 2001; Sadras et al., 2013).

### **Anthocyanins and Flavonols**

The concentration of total anthocyanins in the berries did not vary among treatments at harvest in both vintages in all varieties, so it could be assumed that light conditions were appropriate for anthocyanin biosynthesis in control vines and no improvement arose from bunch light exposure at veraison. Despite in the current study the temperature increase after leaf removal in both years seems not to induce a negative impact on the anthocyanin concentration, its reduction in berries under temperature rise is reported in several papers (Downey et al., 2006; Kliewer and Torres, 1972; Movahed et al., 2016; Mori et al., 2005; Mori et al., 2007).

On these bases, it could not be ruled out that our results may depend by the synergistic effect of higher berry temperature on defoliated vines which may have reduced anthocyanin concentration counterbalancing the supposed enhancement due to light exposure increase.

The multivariate approach applied on the complete anthocyanin concentration data sets allowed the varieties to be differentiated independently of treatments and seasons. The association of Sangiovese and Raboso Piave and their separation from Cabernet Sauvignon and Nero d'Avola is mainly driven by their typical anthocyanin profile, featuring a higher concentration of peonidin 3-glucoside and cyanidin 3-glucoside and a lower concentration of malvidin 3-glucoside in comparison to the other two varieties. In Sangiovese, the effect of veraison defoliation on anthocyanin concentration was stable between the two vintages, causing a clear separation between control and defoliated vines due to the increase in the di-substituted to tri-substituted ratio and this last effect was present also in Raboso Piave.

Cabernet Sauvignon and Nero d'Avola share a similar anthocyanin profile characterized by a high concentration of the three forms of malvidin present in grapevine and low level of di/tri ratio and, according to multivariate approach, showed a general higher stability to treatments and seasons compared to Sangiovese and Raboso Piave in terms of anthocyanins composition. These results seem confirm what recently reported on the higher stability of Cabernet Sauvignon in secondary metabolite composition respect to other black berries varieties (Ortega-Regules et al., 2006) and to Sangiovese. Our outcomes suggest that Raboso Piave could be the most susceptible variety to sunburn among the varieties included in the present study. The increase of di/tri ratio after defoliation in Nero d'Avola, Sangiovese and partially in Raboso Piave cultivars seems to disagree with previous findings referring to both light and temperature increases effects (Mori et al., 2005, Tarara et al, 2008), or with other researches reporting that light exclusion induces an increase of the di/tri ratio compared to control bunches (Downey et al., 2004). It should be considered that in our experimental vineyard, bunches of control vines were naturally shaded and that conditions were not comparable to the one obtained through the light exclusion imposed in the cited research. Moreover, the increase of di-substituted anthocyanins we registered is not in agreement with their supposed lower stability at high temperature due to the chemical degradation hypothesis reported by several authors (Cohen et al,

2012; Mori et al., 2007). Anyway, our biochemical results were supported by other researches in Sangiovese (Pastore et al., 2013) and in Nebbiolo (Guidoni et al., 2008). Despite the total flavonol concentration appeared very variable among the four cultivars in the study, it was very different between control and defoliated vines in all varieties in both vintages. The higher bunch exposure induced by leaf removal in comparison to control berries resulted in an increase of total flavonols in all varieties. Sunlight is known to enhance flavonol accumulation in berries (Downey et al., 2006) and there is a strong positive correlation between illumination and flavonol levels, reflecting their role as UV protectants (Pastore et al., 2013; Price et al., 1995; Spayd et al., 2002). Previous research on Sangiovese showed that, under uniform light conditions, temperature increase caused strong flavonol concentration reduction compared to control, suggesting a negative effect of high temperature on flavonol synthase (Movahed et al., 2016). In our research, the temperature rise was associated with an increase in percentage of bunch exposed to light and in flavonol concentration, revealing that the influence of light is dominant on the synthesis of these compounds compared to the thermal effect, at least under the observed temperature range.

As previously described the total content and pattern of flavonols is highly variable across genotypes and our results confirm that red grape varieties like Sangiovese synthesize mainly di-substituted derivatives like quercetin (Flamini et al., 2013). In control vines, Cabernet Sauvignon and Raboso Piave have similar proportions of myricetin and quercetin, while Nero d'Avola exhibits a high concentration of myricetin. Kaempferol is present in no or low concentration in all the varieties included in this study. The multivariate approach applied on the complete flavonol concentration data sets separated the control from defoliated vines due to the significant increase in the latter, mainly driven by the rise of quercetin which appears the compound more responsive to light, as previously reported by other authors on Tempranillo (Carbonell-Bejerano et al., 2014). In our experimental conditions, this response drives towards a reduction in the differences between the original flavonol profiles of the four varieties.

## Conclusion

In our conditions, where control berries were naturally shaded and subjected to quite high level of temperature which overcome 30° C for several hours, the response of four varieties to veraison defoliation in terms of anthocyanins accumulation remain unclear. We could not exclude that the similar anthocyanin content between treatments in all varieties is caused by the balancing of anthocyanins biosynthesis and degradation induced by the combined effects respectively of light and temperature. The strong increase in flavonol concentration in all varieties under defoliation suggests that the influence of light is dominant on the synthesis of these compounds compared to the thermal effect and that they may represent a marker of berries sun exposure. Furthermore, the stimulation of the synthesis of quercetin, derived from the di-substituted branch of the flavonoids pathway, also triggers the production of cyanidin, suggesting that defoliation may induce, according to genotypes, a specific response at the split-up point of the biosynthesis of di- and tri-substituted flavonoids with consequences on the profile of both anthocyanins and flavonols. Based on the overall results obtained it appears that the relationship between anthocyanin and flavonols and veraison defoliation is very complex and depends on many factors including genotype and the synergistic or antagonistic effect of different levels and extent of both temperature and light intensity experienced by the berries.

## References

- Bergqvist, J., Dokoozlian, N., Ebisuda, N., 2001. Sunlight exposure and temperature effects on berry growth and composition of Cabernet Sauvignon and Grenache in the Central San Joaquin Valley of California. *Am. J. Enol. Vitic.* 52 (1), 1-7.
- Bledsoe, A.M., Kliewer, W.M., Marois, J.J., 1988. Effects of timing and severity of leaf removal on yield and fruit composition of Sauvignon blanc grapevines. *Am. J. Enol. Vitic.* 39, 49-54.
- Chorty, E., Guidoni, S., Ferrandino, A., Novello, V., 2010. Effect of different bunch sunlight exposure levels on ripening and anthocyanin accumulation in Nebbiolo grapes. *Am. J. Enol. Vitic.* 61(1), 23-30.
- Cohen, S.D., Tarara, J.M., Gambetta, G.A., Matthews, M.A., Kennedy, J.A., 2012. Impact of diurnal temperature variation on grape berry development, proanthocyanidin accumulation, and the expression of flavonoid pathway genes. *J. Exp. Bot.* 63(7), 2655-2665.
- Coombe, B. G., 1986. Influence of temperature on composition and quality of grapes. In Symposium on Grapevine Canopy and Vigor Management, XXII IHC 206, 23-36.
- Crippen, D.D., Morrison, J.C., 1986 a. The effect of sun exposure on the compositional development of Cabernet Sauvignon. *Am. J. Enol. Vitic.* 37, 235-242.
- Crippen, D.D., Morrison, J.C., 1986 b. The effect of sun exposure on the phenolic content of Cabernet Sauvignon berries during development. *Am. J. Enol. Vitic.* 37, 243-247.
- Dokoozlian, N.K., Kliewer, W.M., 1996. Influence of light on grape berry growth and composition varies during fruit development. *J. Amer. Soc. Hort. Sci.* 121(5), 869–874.
- Downey, M.O., Dokoozlian, N.K., Krstic, M.P., 2006. Cultural practice and environmental impacts on the flavonoid composition of grapes and wine: a review of recent research. *Am. J. Enol. Vitic.* 57, 257–268.
- Downey, M.O., Harvey, J.S., Robinson, S.P., 2003. Synthesis of flavonols and expression of flavonol synthase genes in the developing grape berries of Shiraz and Chardonnay (*Vitis Vinifera* L.). *Aust. J. Grape Wine Res.* 9, 110-121.
- Downey, M.O., Harvey, J.S., Robinson, S.P., 2004. The effect of bunch shading on berry development and flavonoid accumulation in Shiraz grapes. *Aust. J. Grape Wine Res.* 10, 55-73.
- English, J.T., Thomas, C.S., Marois, J.J., Gubler, W.D., 1989. Microclimates of grapevine canopies associated with leaf removal and control of Botrytis bunch rot. *Phytopathology.* 79, 395- 401.
- Flamini, R., Mattivi, F., Rosso, M.D., Arapitsas, P., Bavaresco, L., 2013. Advanced knowledge of three important classes of grape phenolics: anthocyanins, stilbenes and flavonols. *Int. J. Mol. Sci.* 14(10), 19651-19669.
- Guidoni, S., Ferrandino, A., Novello, V., 2008. Effects of seasonal and agronomical practices on skin anthocyanin profile of Nebbiolo grapes. *Am. J. Enol. Vitic.* 59(1), 22-29.
- Hale, C.R., Buttrose, M.R., 1974. Effect of temperature on ontogeny of berries of *Vitis Vinifera* L., cv. Cabernet Sauvignon. *J. Amer. Soc. Hort. Sci.* 99, 390-394.
- Haselgrove, L., Botting, D., Van Heeswijck, R.V., Høi, P.B., Dry, P.R., Ford, C., Iland, P.G., 2000. Canopy microclimate and berry composition: the effect of bunch exposure on the phenolic composition of *Vitis vinifera* L. cv Shiraz grape berries. *Aust. J. Grape Wine Res.* 6, 141-149.
- Hunter, J.J., De Villiers, O.T., Watts, J.E., 1991. The effect of partial defoliation on quality characteristics of *Vitis vinifera* L. cv Cabernet Sauvignon grapes. II. Skin color, skin sugar, and wine quality. *Am. J. Enol. Vitic.* 42 (1), 13-18.
- Iland, P.O., 1988. Leaf removal effects on fruit composition. In Proceedings of the Second International Symposium for Cool Climate Viticulture and Oenology. RE Smart et al.(eds.). 2, 137-138.

Kliewer, W.M., Lider, L.A., 1968. Influence of bunch exposure to the sun on the composition of Thompson Seedless fruit. *Am. J. Enol. Vitic.* 19, 175-184.

Kliewer, W.M., Torres, R.E., 1972. Effect of controlled day and night temperatures on grape coloration. *Am. J. Enol. Vitic.* 23 (2), 71-77.

Lemut S.M., Trost, K., Sivilotti, P., Arapitsas, P., Vrhovsek, U., 2013. Early versus late leaf removal strategies for Pinot Noir (*Vitis vinifera* L.): effect on colour-related phenolics in young wines following alcoholic fermentation. *J. Sci. Food Agric.* 93(15), 3670-3681.

Lorenz, D. H., Eichhorn, K. W., Bleiholder, H., Klose, R., Meier, U., Weber, E., 1995. Growth stages of the grapevine: phenological growth stages of the grapevine (*Vitis vinifera* L. ssp. *vinifera*)—codes and descriptions according to the extended BBCH scale. *Aust. J. Grape Wine Res.* 1(2), 100-103.

Mattivi, F., Guzzon, R., Vrhovsek, U., Stefanini, M., Velasco, R., 2006. Metabolite profiling of grape: flavonols and anthocyanins. *J. Agric. Food Chem.* 54, 7692–7702.

Matus, J.T., Loyola R., Vega A., Peña-Neira, A., Bordeu E., Arce-Johnson P., Alcalde J.A., 2009. Post-veraison sunlight exposure induces MYB-mediated transcriptional regulation of anthocyanin and flavonol synthesis in berry skins of *Vitis vinifera*. *J. Exp. Bot.* 60, 853-867.

Mori, K., Goto-Yamamoto, N., Kitayama, M., Hashizume, K., 2007. Loss of anthocyanins in redwine grape under high temperature. *J. Exp. Bot.* 58(8), 1935-1945.

Mori, K., Sugaya, S., Gemma, H., 2005. Decreased anthocyanin biosynthesis in grape berries grown under elevated night temperature condition. *Sci. Hortic.* 105, 319–330.

Movahed, N., Pastore, C., Cellini, A., Allegro, G., Valentini, G., Zenoni, S., Cavallini, E., D'Inca, E., Tornielli, G.B., Filippetti, I., 2016. The grapevine VviPrx31 peroxidase as a candidate gene involved in anthocyanin degradation in ripening berries under high temperature. *J. Plant Research*, 129(3), 513-526.

Pastore, C., Zenoni, S., Fasoli, M., Pezzotti, M., Tornielli, G.B., Filippetti, I., 2013. Selective defoliation affects plant growth, fruit transcriptional ripening program and flavonoid metabolism in grapevine. *BMC Plant Biology*. DOI: 10.1186/1471-2229-13-30.

Percival, D.C., Fisher, K.H., Sullivan, J.A., 1994. Use of fruit zone leaf removal with *Vitis vinifera* L. cv Riesling grapevines. II. Effects on fruit composition, yield, and occurrence of bunch rot (*Botrytis cinerea* Pers.). *Am. J. Enol. Vitic.* 45, 33-139.

Pereira, G.E., Gaudillere, J.P., Pieri P., Hilbert, G., Maucourt, M., Deborde, C., Moing, A., Roil, D., 2006. Microclimate influence on mineral and metabolic profiles of grape berries. *J. Agric. Food Chem.* 54, 6765-6775.

Poni, S., Intrieri, C., Silvestroni, O., 1994. Interactions of leaf age, fruiting, and exogenous cytokinins in Sangiovese grapevines under nonirrigated conditions. I. Gas exchange. *Am. J. Enol. Vitic.* 45, 71-78.

Price, S.F., Breen, P.J., Valladao, M., Watson, B.T., 1995. Bunch sun exposure and Quercetin in grapes and wine. *Am. J. Enol. Vitic.* 46, 187-194.

Ristic, R., Downey, M.O., Iland, P.G., Bindon, K., Francis, I.L., Herderich, M., Robinson, S.P., 2007. Exclusion of sunlight from Shiraz grapes alters wine colour, tannin and sensory properties. *Aust. J. Grape Wine Res.* 13(2), 53-65.

Rustioni, L., Milani, C., Parisi, S., & Failla, O., 2015. Chlorophyll role in berry sunburn symptoms studied in different grape (*Vitis vinifera* L.) cultivars. *Sci. Hort.*, 185, 145-150.

Sadras, V.O., Petrie, P.R., Moran, M.A., 2013. Effects of elevated temperature in grapevine. II juice pH, titratable acidity and wine sensory attributes. *Aust. J. Grape Wine Res.* 19(1), 107-115.

Spayd, S.E., Tarara, J.M., Mee, D.L., Ferguson, J.C., 2002. Separation of sunlight and temperature effects on the composition of *Vitis vinifera* cv. Merlot berries. *Am. J. Enol. Vitic.* 53, 171-182.

Stapleton, J.J., R. Stanley Grant, 1992. Leaf removal for nonchemical control of the summer bunch rot complex of wine grapes in the San Joaquin Valley. *Plant disease*. 76.2, 205-208.

Tarara, J.M., Lee, J., Spayd, S.E., Scagel, C.F., 2008. Berry temperature and solar radiation alter acylation, proportion, and concentration of anthocyanin in merlot grapes. *Am. J. Enol. Vitic.* 59(3), 235–247.

Zoecklein, B. W., Wolf, T. K., Duncan, N. W., Judge, J. M., Cook, M. K., 1992. Effects of fruit zone leaf removal on yield, fruit composition, and fruit rot incidence of Chardonnay and white Riesling (*Vitis vinifera* L.) grapes. *Am. J. Enol. Vitic.* 43.2, 139-148.